

## ***In vitro* Free Radical Scavenging, Anti-Hyaluronidase and Anti-Elastase Potential of Flaxseed Lignans Concentrate**

Jayarama Naik N.<sup>1</sup> and Basavaraj Madhusudhan<sup>1,2\*</sup>

<sup>1</sup>Department of Food Technology, Davangere University, Davangere – 577 007, Karnataka, India

<sup>2</sup>Research Center for Nanoscience and Technology, Department of Biochemistry and Food Technology, Davangere University, Shivagangothri, Davangere- 577007, Karnataka, India

\*Corresponding Author E-mail: [prof.basavarajmadhusudhan@gmail.com](mailto:prof.basavarajmadhusudhan@gmail.com)

Received: 2.11.2018 | Revised: 7.12.2018 | Accepted: 14.12.2018

### **ABSTRACT**

*The present study was carried out to study the efficiency of flaxseed lignans concentrate (FLC) on hyaluronidase and elastase enzymes inhibition activity. The DPPH and FRAP Assay were carried out using butylated hydroxytoluene and Iron (II) sulfate as standard molecules. The DPPH radical scavenging ability of FLC had the IC<sub>50</sub> value of 486 µg ml<sup>-1</sup> and FRAP assay of FLC had IC<sub>50</sub> value of 74.12 µg ml<sup>-1</sup>. Furthermore, oleanolic acid was used as reference for enzyme inhibitory assays. The IC<sub>50</sub> value for hyaluronidase inhibitory activity of FLC was 10.75 µg ml<sup>-1</sup> whereas oleanolic acid has IC<sub>50</sub> value of 9.93 µg ml<sup>-1</sup>. FLC showed elastase inhibition with IC<sub>50</sub> value of 51.44 µg ml<sup>-1</sup> whereas standard molecule oleanolic acid had IC<sub>50</sub> value of 41.72 µg ml<sup>-1</sup>. It can be summarized from the above observations that flaxseed being a rich source of lignans and rich anti-oxidant properties, the flaxseed lignans concentrate has the potential similar to oleanolic acid and may be utilized as anti-wrinkle agent in cosmetic products.*

**Key words:** Flaxseed, Anti-wrinkle, Anti-aging, Lignans, Enzymes inhibition.

### **INTRODUCTION**

When the skin repeatedly exposed to solar radiation it produces excessive free radicals these free radicals causes oxidative stresses, inflammation and destruction of antioxidant system as a result cell membranes, lipids, proteins and DNA get damaged<sup>1</sup>. Increased hyaluronidase and elastase enzymes in skin tissues are responsible for degradation of dermal network which further leads to skin wrinkling and aging. Thus, inhibition of these enzymes is essential in anti-aging prevention<sup>2,3</sup>. Skin contains hyaluronic acid

and elastin to maintain elasticity and structure of extracellular matrix of connective tissues. However, the enzyme elastase can degrade it<sup>4</sup>. As a consequence, these substances declines sharply during mature and pre-mature aging<sup>5,6</sup>. Loss of hyaluronic acid and elastin leads to loss of elasticity and water retention capacity of skin<sup>7</sup>. Therefore, to eliminate free radicals from the body through antioxidant defense system and sustaining antioxidant homeostasis always there is an opportunity for new natural antioxidants to prevent skin aging<sup>8</sup>.

**Cite this article:** Naik, J.N. and Madhusudhan, B., *In vitro* Free Radical Scavenging, Anti-Hyaluronidase and Anti-Elastase Potential of Flaxseed Lignans Concentrate, *Int. J. Pure App. Biosci.* 6(6): 172-179 (2018). doi: <http://dx.doi.org/10.18782/2320-7051.7059>

*Linum usitatissimum* L. (family: Linaceae), commonly well-known as 'Flaxseed' or 'Linseed'. Since ages it has been grown for oil and fibre purpose<sup>9</sup>. Major flaxseed growing countries are Canada, China, United States and India<sup>10</sup>. Flaxseed represent richest source of phenolic antioxidant called secoisolariciresinol diglucoside (SDG)<sup>11,12</sup>. SDG without glucose molecule in its structure called as Secoisolariciresinol (SECO) which is insoluble in water<sup>13</sup>. Flaxseed lignans SDG, SECO, ED and EL are found to be equal or somewhat more potent than BHT, vitamin E<sup>14</sup>. Thus, they could have commercial potential as an alternative to these antioxidants<sup>15</sup>. However, so far very limited research attention has been given on flaxseed lignans as an anti-aging agent. Hence flaxseed lignans concentrate was prepared and tested its potential as an ingredient to relieve the skin wrinkling and aging symptoms.

## MATERIAL AND METHODS

### 1. Extraction and preparation of flaxseed lignans concentrate

Briefly, the flaxseed sample (10 kg) was washed and subjected for a dehulling process to obtain hull fraction of flaxseed using Kisan Krishi Yantra Udyog dehuller at Grain Science and Technology Department, CFTRI, Mysore, India. Flaxseeds hull fraction was taken, defatted by extracting with n-hexane. The lignans concentrate was prepared from defatted hull fraction of flaxseed. About 10 g of defatted hull fraction was ground and sieved, mixed with 400 ml of distilled water followed by 500 ml of 2 M aqueous sodium hydroxide. The contents were incubated for 1 h at 20 °C on a shaking water bath. The hydrolysate was then acidified with dilute sulphuric acid to pH 3.0 and then centrifuged at 5000 rpm for 10 min. The supernatant was centrifuged rapidly to a clear liquid phase and pooled together. The liquid phase (600ml) was mixed with 95% aq. ethanol (900ml), left at room temperature for at least 10 min and total volume was divided into four equal volumes in centrifuge tubes and again centrifuged at 10000 rpm for 5 min to precipitate. Water

soluble polysaccharide and proteins were removed carefully. Ethanol extract was evaporated by using rotary evaporator at 40 °C to obtain FLC. The lyophilized FLC was stored until further analysis<sup>16</sup>.

### 2. DPPH radical scavenging assay

The stable free radical-scavenging activity was determined by the DPPH assay. In this method, the bleaching rate of a stable free radical, DPPH is monitored at a characteristic wavelength in the presence of the various phytochemicals used in the study. In its radical form, DPPH absorbs at 517 nm, but upon reduction by an antioxidant or a radical species, its absorption decreases. Different concentrations of FLC @ 100, 200, 300, 400 and 500 µg (each in three replicates) were added, at an equal volume, to methanolic solution of DPPH (100 M). After 15 min at room temperature, the absorbance was recorded at 517 nm while IC<sub>50</sub> values denote the concentration of sample, which is required to scavenge 50% of DPPH free radicals<sup>17</sup>.

### 3. FRAP assay

The FRAP reagent was prepared from acetate buffer (pH 3.6), 10 mmol 2,4,6-tripyridyl-s-triazine (TPTZ) solution in 40 mmol HCl and 20 mmol iron (III) chloride solution in proportions of 10:1:1 (v/v/v), respectively. 100 µl, 200 µl and 300 µl of FLC (each in three replicates) were added to 1.5 mL of the FRAP reagent. The absorbance of the reaction mixture was then recorded at 593 nm after 4 min.

The standard curve was constructed using iron (II) sulfate solution (100–2000 g mL<sup>-1</sup>), and the results were expressed as g mL<sup>-1</sup> Fe (II) of various concentrations of FLC used in this study<sup>18</sup>.

### 4. Elastase inhibitory activity assay

Elastase inhibitory activity was measured by modified method of Sigma. A mixture of 10 µl to 50 µl of FLC, 5 µl elastase from porcine pancreas [Sigma 45124, USA] (0.5 mU mL<sup>-1</sup> in the cool aquades) and 125 µL Tris buffer was pre incubated for 15 min at 25°C. Mixed solution was added N-Sucanyl-Ala-Ala-Ala-p-Nitroanilide substrate<sup>19</sup> and then incubated for 15 min at 25°C.

Absorbance was measured at 410 nm wavelength.

$$\% \text{ Elastase inhibition} = (1-B/A) \times 100\%$$

A = sample absorbance

B = control absorbance

## 5. Hyaluronidase Inhibitory Activity

### Assay

Hyaluronidase inhibitory of activity was measured by modified method of Sigma Aldrich. A 5 to 25  $\mu\text{L}$  of FLC solution and 3  $\mu\text{L}$  hyaluronidase from bovine testes type I-S [Sigma H3506, USA] was pre-incubated for 10 min at 37 °C and then added 12  $\mu\text{L}$  phosphate buffer (300 mM, pH 5.35) for 10 min at 37 °C. Afterward 10  $\mu\text{L}$  hyaluronic acid substrate [Sigma H5542, USA] was added and incubated for 45 min at 37 °C. Decomposition reaction of hyaluronic acid was stopped by adding 100  $\mu\text{L}$  acidic albumin acid. Mixed solution incubated at room temperature for 10 min, then absorbance was measured at 600 nm wavelength<sup>20</sup>.

Quantification of inhibition activity is expressed as:

$$\% \text{ Hyaluronidase inhibition} = (1-B/A) \times 100\%$$

A = sample absorbance

B = control absorbance

## STATISTICAL ANALYSIS

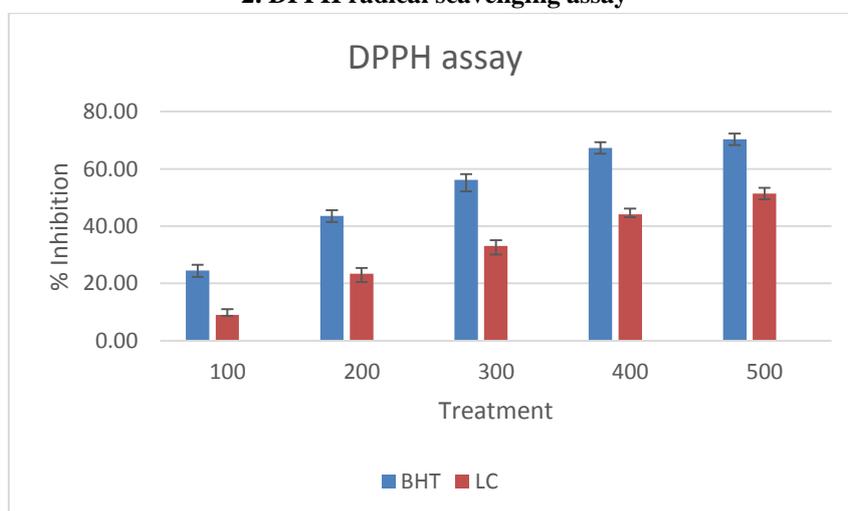
All the experiments were repeated at least three times with three replications for each treatments. Therefore, the data represent the means and standard errors (Mean  $\pm$  SD) and were calculated using Microsoft Excel 2010 software.

## RESULTS

### 1. Extraction of Flaxseed lignans

Several methods have been reported by the earlier researchers for the extraction of lignans. The highest SDG content could be extracted by the method of<sup>21</sup>. On dry matter basis, different samples of flaxseeds varied considerably in their content of SDG (11.9–25.9  $\text{mg g}^{-1}$ ) along with p-coumaric acid glucoside (1.2–8.5  $\text{mg g}^{-1}$ ), and ferulic acid glucoside (1.6–5.0  $\text{mg g}^{-1}$ ). In our study, the SDG content of LC was found to be 23.28  $\text{mg g}^{-1}$ .

### 2. DPPH radical scavenging assay

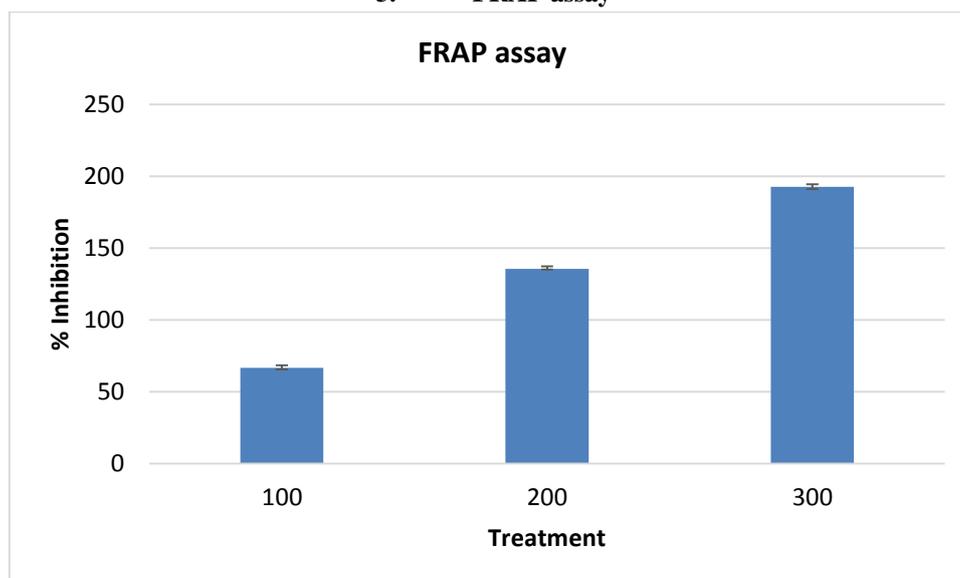


**Fig. 5: 1-diphenyl-2-picrylhydrazyl assay of different formulation Each bars point is the Mean  $\pm$  SD of three replicates from three independent experiments.**

The results of the antioxidant, free radical scavenging of lignans concentrate was presented in the Figure 5. The DPPH radical

scavenging ability of lignans concentrate has the  $\text{IC}_{50}$  value of 486  $\mu\text{g ml}^{-1}$  whereas BHT has 267  $\mu\text{g ml}^{-1}$ .

## 3. FRAP assay

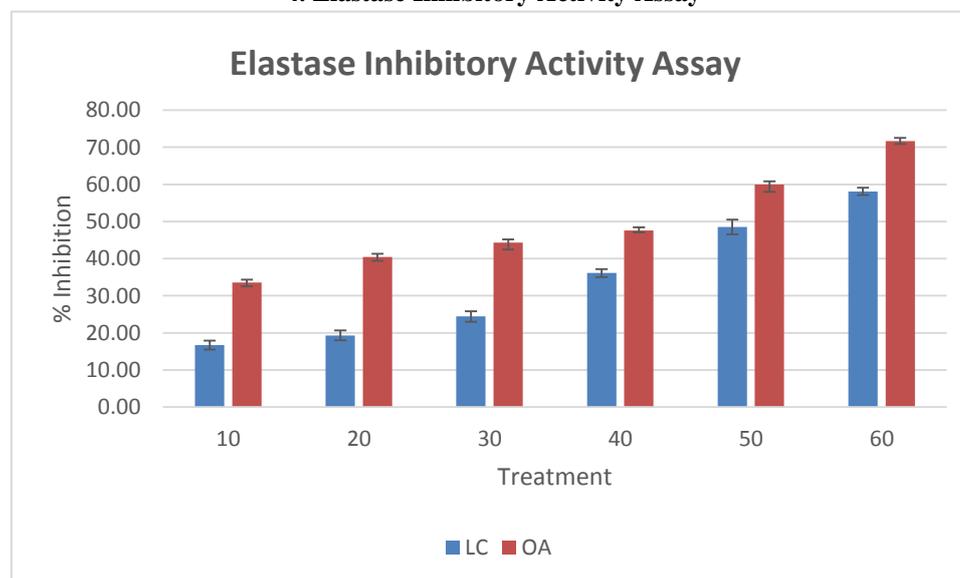


**Fig. 6:** Ferric reducing antioxidant power assay of different concentration Each bars point is the Mean  $\pm$ SD of three replicates from three independent experiments

The FRAP assay was carried out according to the procedure Benzie and Strain 1996. The standard curve was constructed using Iron (II) sulfate solution ( $\mu\text{g ml}^{-1}$ ). The results were

expressed as  $\mu\text{g ml}^{-1}$  Fe (II) of lignans concentrate used in study. Lignans concentrate has  $\text{IC}_{50}$  value of  $74.12 \mu\text{g ml}^{-1}$ .

## 4. Elastase Inhibitory Activity Assay

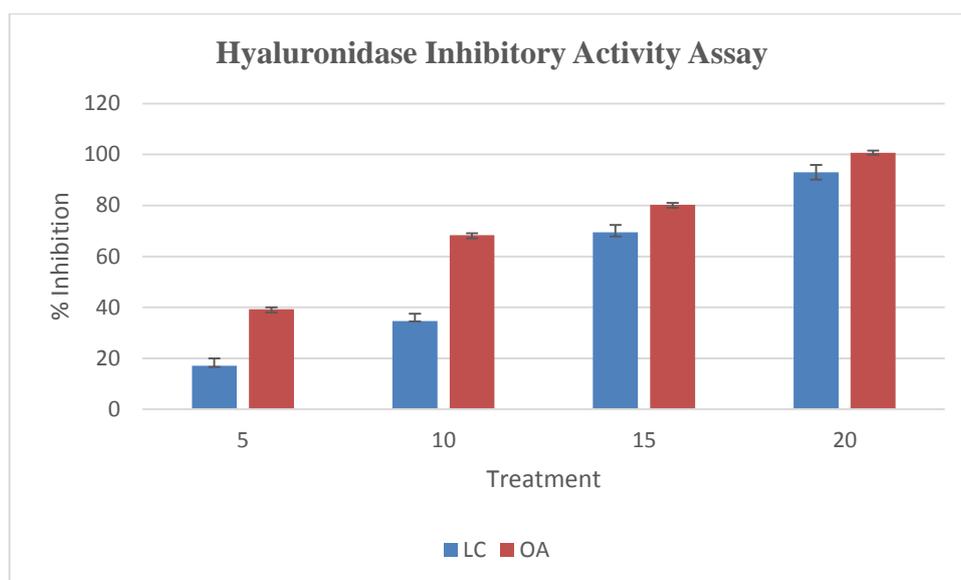


**Fig. 7:** Elastase inhibitory activity of Lignans Concentrate and Oleanolic Acid Each bars point is the Mean  $\pm$ SD of three replicates from three independent experiments

The elastase inhibitory activity were measured. Lignans concentrate showed elastase inhibition with  $\text{IC}_{50}$  value of  $51.44 \mu\text{g ml}^{-1}$  whereas

standard molecule oleanolic acid has  $\text{IC}_{50}$  value of  $41.72 \mu\text{g ml}^{-1}$ .

### 5. Hyaluronidase Inhibitory Activity Assay



**Fig. 8: Hyaluronidase inhibitory activity of Lignans Concentrate and Oleanolic Acid Each bars point is the Mean  $\pm$  SD of three replicates from three independent experiments**

Hyaluronidase was assayed by a highly sensitive spectroscopic method, based on precipitation of hyaluronic acid with cetylpyridinium chloride, which is used for high throughput screening for hyaluronidase inhibitors. The  $IC_{50}$  value for hyaluronidase inhibitory activity of lignans concentrate was  $10.75 \mu\text{g ml}^{-1}$  whereas oleanolic acid had  $IC_{50}$  value of  $9.93 \mu\text{g ml}^{-1}$ .

#### DISCUSSION

The DPPH method is commonly used worldwide for *in vitro* quantification of free radical scavenging activity<sup>22</sup>. In the present study, the investigation of total antioxidant capacity was measured as the capacity of the flaxseed lignans concentrate to scavenge stable organic free radicals with colour development, which gave the absorbance maxima at 517nm. As this method is sensitive to light, oxygen, pH and type of solvent used<sup>23</sup> extra care was taken to overcome these. It has been proved that phenolic, flavonoids compounds and other secondary metabolites present in the plants are mainly responsible for antioxidant activity<sup>24</sup>. From the above results, it can be confirmed that lignans, flavanoids and polyphenolics proved their higher efficiency as an antioxidant. For further confirmation of

antioxidant potential of Flaxseed lignans concentrate FRAP assay was carried out. The phenolic phytochemicals exhibits oxidation-reduction properties, which play a crucial role in determining the total antioxidant properties<sup>25</sup>. In the present study, the reducing ability of FLC was strongly correlated with the percent inhibition by standard molecule. It has been established that elastin, collagen and hyaluronic acid are the main components of skin which have an important role in maintaining skin structure and hydration<sup>26</sup>. Moreover, elastin is a fibrose protein that compose 2–4% of the ECM and involved in the hydration of the skin<sup>29</sup>. On the other hand, collagenase, hyaluronidase and elastase cause repetitive collagen fibers breakdown and responsible for structural defect in dermis and wrinkle development<sup>27</sup>. Collagenase and elastase contribute in production and degradation of fibers, which is also influenced by free radical oxidative stress<sup>28</sup>. The enzyme elastase is responsible for increased tissue permeability, inflammation progress and delayed wound healing<sup>30</sup>. Moreover, elastase is also the key enzyme that attacks all the major connective tissue matrix protein<sup>31</sup>. While, the enzyme hyaluronidase selectively degrade hyaluronic acid by lowering its viscosity and

increasing the permeability<sup>32</sup>. Which is an acidic structural polysaccharide found exclusively in the extracellular matrix (ECM). The HPLC analysis of flaxseed lignans concentrate revealed that it includes major fraction as SDG along with p-coumaric acid glucoside and ferulic acid glucoside<sup>21</sup>. Moreover, a smaller quantities of other type lignans such as matairesinol, isolariciresinol, lariciresinol and pinoresinol have also been identified in the flaxseed<sup>9</sup>. Further, it has been proved that phenolic, flavonoids compounds and other secondary metabolites present in the plants are known to inhibit extracellular matrix degradative enzymes. Hyaluronidase inhibitors are thus potent regulators that maintain hyaluronic acid homeostasis and they might serve as anti-inflammatory, anti-aging, antimicrobial, anti-cancer and antitoxin and contraceptive agents<sup>33</sup>.

### CONCLUSION

From the present study, it was found that lignans extracted from defatted flaxseed hull potentially inhibited the hyaluronidase and elastase enzyme activity. It also showed free radical scavenging activity. Thus, it can be concluded that Flaxseed Lignans may have potential role on skin care, which has very good economic potential and being cultivated in more than 50 countries. In addition to SDG, smaller quantities of other type lignans such as matairesinol, isolariciresinol, lariciresinol and pinoresinol have also been identified in the flaxseed. SDG is converted into mammalian lignans, enterodiol (ED) and enterolactone (EL) by colon bacteria can be explored further for its use in cosmetics for its enzyme inhibition activity.

### Acknowledgements

Authors are thankful DST-SERB sponsored project Grant No SB/EMEQ-429/2014 and for providing Senior Research Fellowship. Thanks are also to Davangere University, Karnataka, for facilities and encouragement.

### Conflict of interest

The authors declare that there are no conflicts of interest.

### REFERENCES

1. Kim, H., Yun, J., Lee, J., Hong, H., Jeong, J. and Kim, E., SUMO1 attenuates stress-induced ROS generation by inhibiting NADPH oxidase 2, *Biochem. Biophys. Res. Commun.*, **410(3)**: 555–62 (2011).
2. Labat-Robertm, J., Fourtanier, A., Boyer-Lafargue, B. and Robert, L., Age dependent increase of elastase type protease activity in mouse skin: effect of UV-irradiation, *J. Photochem. Photobiol. B. Biol.*, **57**: 113–118 (2000).
3. Yamamoto, Y. J., Role of active oxygen species and antioxidants in Photoaging, *J. Dermatol. Sci.*, **27**: S1–S4 (2001).
4. Thing, T. S., Hili, P. and Naughton, D. P., Anti-collagenase anti-elastase and antioxidant activities of extracts from 21 plants, *BMC Compl. Alter. Med.*, **9(27)**: 1–11 (2000).
5. Manuskiatti, W. and Maibach, H., Hyaluronic acid and skin wound healing and aging. *Int. J. Dermatol.*, **35**: 539–541 (1996).
6. Reed, R. K., Saito, M. and Qiu, G., Hyaluronan in the rat with special reference to the skin, *Acta. Physiol. Scand.*, **134**: 405–411 (1988).
7. Ryu, A., Naru, E. and Arakane, K., Cross-linking of collagen by singlet oxygen generated with UV-A, *Chem. Pharm. Bull.*, **45**: 1243–1247 (1997).
8. Wang, K. H., Lin, R. D., Hsu, F. L., Huang, Y. H., Chang, H. C., Huang, C. Y. and Lee, M. H., Cosmetic applications of selected traditional Chinese herbal medicines, *J. Ethnopharmacol.*, **106**: 353–359 (2006).
9. Madhusudhan, B., Potential benefits of flaxseed in health and disease – a perspective, *Agriculturae. Conspectus. Scientificus*, **74**: 67-72 (2009).
10. Rubilar, M., Gutiérrez, C., Verdugo, M., Shene, C. and Sineiro, J., Flaxseed as a source of functional ingredients, *J. of Soil Sci. and plant nutri.*, **10**: 373 – 377 (2010).
11. Bambagiotti-alberti, M., Coran, S. A., Ghiara, C., Moneti, G. and Raffaelli, A.,

- Investigation of mammalian lignan precursors in flax seed: first evidence of secoisolariciresinol diglucoside in two isomeric forms by liquid chromatography/mass spectrometry, *Rap. Commun. in Mass Spec.*, **8**: 929-932 (1994).
12. Cardoso carraro, J. C., Inês de souza dantas, M., Rocha espescht, A. C., Duarte martino, H. S. and Rocha ribeiro, S. M., Flaxseed and Human Health: Reviewing Benefits and Adverse Effects, *Food Rev. Int.*, **28**: 203-230 (2012).
  13. Mazur, W., Fotsis, T., Wähälä, K., Ojala, S., Salakka, A. and Adlercreutz, H., Isotope dilution gas chromatographic-mass spectrometric method for the determination of isoflavonoids, coumestrol, and lignans in food samples, *Anal. Biochem.*, **233**: 169-180 (1996).
  14. Prasad, K., Antioxidant activity of secoisolariciresinol diglucoside-derived metabolites, secoisolariciresinol, enterodiol, and enterolactone, *Int. J. of Ang.*, **9**: 220–225 (2000).
  15. Hu, C., Yuan, Y. V. and Kitts, D. D., Antioxidant activities of the flaxseed lignan secoisolariciresinol diglucoside, its aglycone secoisolariciresinol and the mammalian lignans enterodiol and enterolactone *in vitro*, *Food and Chem. Toxic.*, **45**: 2219–2227 (2007).
  16. Ramsay, A., Fliniaux, O., Quéro, A., Molinié, R., Demailly, H., Hano, C., Paetz, C., Roscher, A., Grand, E., Kovensky, J. and Schneider, B., Kinetics of the incorporation of the main phenolic compounds into the lignan macromolecule during flaxseed development, *Food chem.*, **217**: 1-8 (2017).
  17. Jaberiana, H., Piri, K., and Nazari, J., Phytochemical composition and *in vitro* antimicrobial and antioxidant activities of some medicinal plants, *Food Chem.*, **136**: 237–44 (2013).
  18. Benzie, I. F. F. and Strain, J. J., The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: the FRAP assay, *Anal. Biochem.*, **239**: 70–6 (1996).
  19. Thing, T. S., Hili, P. and Naughton, D. P., Anti-collagenase, antielastase and antioxidant activities of extracts from 21 plants, *BMC Compl. Alter. Med.*, **9(27)**: 1–11 (2009).
  20. Tung, J. S., Mark, G. E. and Hollis, G. F., A microplate assay for hyaluronidase and hyaluronidase inhibitors, *Anal. Biochem.*, **223**: 149–52 (1994).
  21. Eliasson, C., Kamal-Eldin, A., Andersson, R. and Aman, P., High-performance liquid chromatographic analysis of secoisolariciresinoldi glucoside and hydroxycinnamic acid glucosides in flaxseed by alkaline extraction. *J. of Chrom.*, **A1012**: 151 -159 (2003).
  22. Jagtap, U. B., Panaskar, S. N. and Bapat, V. A., Evaluation of antioxidant capacity and phenol content in jackfruit (*Artocarpus heterophyllus* Lam.) fruit pulp, *Plant Foods Hum. Nutr.*, **65**: 99–104 (2010).
  23. Ozcelik, C., Lee, J. H. and Min, D. B., Effects of light, oxygen and pH on the absorbance of 2,2 diphenyl 1 picrylhydrazyl, *J. Food Sci.*, **68**: 487–90 (2003).
  24. Balsundram, N., Sudram, K. and Samman, S., Phenolic compounds in plants and agro industrial by products: antioxidant activity, occurrence and potential uses, *Food Che.*, **99**: 191–203 (2006).
  25. Rice-Evans, C. A., Miller, N. T., Paganga, G., Antioxidant properties of phenolic compounds, *Tre, in Plant Sci.*, **4**: 304–9 (1997).
  26. Hooda, R., Anti-wrinkle herbal drugs-an update, *J. Pharmacog. Phytochem.*, **4(4)**: 277–8, 2015.
  27. Shehada, A., A review on natural bioactive compounds as potential anti-wrinkle agents, *World J. Pharm.*, **3**: 528– 44 (2014).
  28. Mukherjee, P., Maity, N., Nema, N., Sarkar, B., Bioactive compounds from natural resources against skin aging, *Phytomed.*, **19**: 64–73 (2011).

29. Jenkins, G., Molecular mechanisms of skin ageing, *Mech. Ageing Dev.*, **123**: 801–810 (2002).
30. Siedle, B., Hrenn, A. and Merfort, I., Natural compounds as inhibitors of human neutrophil elastase, *Planta Med.*, **73**: 401–20 (2007).
31. Azmi, N., Hashim, P., Hashim, D. M., Halimoon, N. and Nik Majid, N. M., Anti-elastase, anti-tyrosinase and matrixmetalloproteinase-1 inhibitory activity of earthworm extracts as potential new anti-aging agent, *Asian Pac. J. Trop. Biomed.*, **4(1)**: 348–52 (2014).
32. Girish, K., Kemparaju, K., Nagaraju, S., Vishwanath, B., Hyaluronidase inhibitors: A biological and therapeutic perspective, *Curr. Med. Chem.* **16**: 2261–88 (2009).
33. Girish, K., Kemparaju, K., The Magic Glue Hyaluronan its Eraser Hyaluronidase: A biological overview, *Sci.*, **80**: 1921–43 (2007).