

## An Overview: Mechanism Involved in Bio-Priming Mediated Plant Growth Promotion

Sukanya V.<sup>1</sup>, R. M. Patel<sup>2</sup>, K. P. Suthar<sup>1\*</sup> and D. Singh<sup>1</sup>

<sup>1</sup>Department of Plant Molecular Biology and Biotechnology, ASPEE College of Horticulture and Forestry, Navsari Agricultural University, Navsari, Gujarat, India 396 450

<sup>2</sup>ASPEE SHAKILAM Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India 395 007

\*Corresponding Author E-mail: [kpsmolbio@gmail.com](mailto:kpsmolbio@gmail.com)

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### ABSTRACT

*Application of plant growth promoting rhizobacteria in agriculture is a need of time for sustainable agriculture. Bio-priming is a treatment of seeds with beneficial microorganism under controlled hydration which enhances the preparatory processes prior to germination without the emergence of the radicle. It improves the seedling vigor, germination percentage, speed of germination, growth and development. Bio-priming agents directly release growth promoting hormones viz., indole acetic acid, gibberellin, and cytokinin or stimulate their production in the plant, they also improve the availability of minerals viz., nitrogen, phosphorous, potassium, and Iron whereby enhance the plant growth and development. Bio-priming affects plant by different indirect means viz., phytohormones production stimulation, modulation in different secondary metabolites, alteration in gene expression, tolerance to abiotic and biotic stress. The PGPR bio-priming enhances production of soluble protein, soluble sugar, phenolic acid, salicylic acid and plant growth hormones. Further, efficient mitochondrial development by augmenting energy metabolism is a specialty of bio-primed seeds. The bio-priming also found to induce early DNA and protein synthesis. Bio-priming reported to enhance the expression of RuBisCO and chl a/b, expansins,  $\beta$ -tubulin and GST genes which at the end leads to have better carboxylation capacity and efficient photosynthesis whereby stimulate early germination process and vigorous plant growth. Current biochemical and molecular level understanding confirms the beneficial effect of bio-priming on seed germination, plant stand establishment, growth and development.*

**Key words:** Bio-priming, PGPR, Physiology, Biochemical, Gene expression

### INTRODUCTION

The agriculture sector contributes 7.8 percent to Indian economy which is much higher than the world average i.e., 6.1 percent<sup>1</sup>. Increasing population and awareness of health leads to huge pressure for quality adequate foods at a correct time and that should be free from an unacceptable level of chemicals. To

accomplish this goal, every country switched over to organic farming or sustainable farming. Good quality seeds with rapid germination, producing synchronized vigorous seedlings, higher yield potential and productivity are very important factors for agriculture.

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Seed is preferred technique of agriculture scientist for disseminating any technology as it is easy means of adaptation and cost-effective.

Secretion of nutrients by plant roots enhances the abundance of microorganisms in the rhizosphere<sup>2</sup>. Plant and rhizospheric microorganism have a close association, some are beneficial and others are detrimental. Beneficial microorganisms have the ability to live within or in the vicinity of plant roots and promote plant growth and development are known as plant growth promoting rhizobacteria (PGPR)<sup>3,4</sup>. The direct and indirect effects of PGPR on plant growth and development are noticeable<sup>5,6</sup>. Besides the promotion of plant growth, it possesses the ability to suppress soil born pathogens by producing antibiotics<sup>7</sup> and by induced systemic resistance<sup>8,9</sup>. It also strengthens plants to tolerate under abiotic stress conditions like salt and drought<sup>10,11,12</sup>, chilling stress<sup>13</sup> etc. Further, the PGPR plays an important role in improving and maintaining soil structure and bioremediation of contaminated soil<sup>14</sup>. The PGPR based on the application in agriculture are classified into various groups (Table 1).

Application of PGPR in the agriculture is a potential alternative to chemical-based agriculture which had severely destroyed the agro-ecosystem<sup>34</sup>. The first biological product based on *Bacillus subtilis* was bacteriological fertilizer for inoculation of cereals, marketed in 1897 under the proprietary name “Alinit” by Farbenfabriken<sup>35</sup>. For a sustainable agriculture, symbiotic PGPR (*Rhizobium* and *Frankia*) and non-symbiotic PGPR (*Azotobacter*, *Azospirillum*, *Bacillus* and *Klebsiella*) are commercially applied to enhance yield and reduce the use of harmful agrochemicals<sup>36</sup>. Survival of PGPR on seed and in soil mainly depends on the method of application and competency of the introduced bacteria with other bacteria present in the rhizosphere. Augmentation of PGPR into plants can be achieved by different methodologies including direct soil application, root dipping method, seed coat

pelleting and seed priming. Direct application of inoculums into the soil is advisable when the plant tissue contains some antagonist microbes or pesticidal compounds<sup>37</sup>. It is a simple and easy method but the requirement of large inoculums make it very costly further, it needs special care during transportation and after field application<sup>38</sup>. The root dipping method is mainly focused for biocontrol but it requires preparation of plant nurseries which is not cost effective for some plants<sup>39,40,41</sup>.

Seed pelleting hinders gaseous exchange whereby reduce nitrogen fixation in leguminous seeds so seed treatment is considering as a most effective alternative technique as it required only small dose of microbial inoculum with high efficiency<sup>42</sup>. Seed priming is a modern seed treatment method involves soaking of seeds in a solution of a specific priming agent with restricted water availability under controlled conditions followed by drying of seeds into its original weight that initiates preparatory germination related process<sup>43</sup> and maintaining the seeds in phase II stage of imbibition by extending lag phase of germination<sup>44</sup>. Different seed priming methods are available which are depending on the substances used named as hydro priming, halo priming, osmopriming, hormonal priming and bio-priming. Hydro-priming means seed are soaking in water<sup>45</sup>. Inorganic salts like NaCl, KNO<sub>3</sub>, CaCl<sub>2</sub> and CaSO<sub>4</sub> etc. are used in halo priming<sup>46,47</sup>. The optimum concentration of plant growth hormones like auxin, gibberellin, abscisic acid and ethylene have been used in hormonal priming<sup>48,49,50</sup>. The osmopriming means priming with osmoticants like sugar, mannitol, polyethylene glycol (PEG) etc.<sup>51,52,53</sup> and bio-priming means the use of beneficial microorganisms<sup>54,55</sup>. This review mainly emphasized on physiological, biochemical and molecular changes in seedling of PGPR bio-primed seeds and how these changes are influencing germination, seedling vigor, growth, development and productivity of plants.

## BIO-PRIMING

Bio-priming is an emerging trend aimed to improve seed quality, seedling vigor, productivity and resistance to biotic and abiotic stress by reducing the use of chemical inputs for sustainable agriculture. Bio-priming is a treatment of seed with beneficial microorganism under controlled hydration which enhances the preparatory processes prior to germination without the emergence of the radicle. It is associated with an increase in hydrolytic enzyme activities, reactive oxygen species (ROS) detoxifying enzymes activities and alteration in internal plant hormone levels, and also a differential expression of genes in plants that contributes the enhanced plant growth and resistance against biotic and abiotic stress. Innovative research studies at biochemical, proteomics and transcriptome levels are necessary to understand the role of bio-priming with PGPRs in phyto-stimulation and nutrient enhancement.

### The direct effect of Bio-priming

Plant growth promoting rhizobacteria in primed seeds are colonizing the root surface and competing with other microorganisms in the rhizosphere. Bio-priming is directly involved in the enhancement of plant growth by the secretion of compounds and mineral solubilization. Nitrogen is an important constituent of amino acid, chlorophyll and other structural components of plants<sup>56</sup>. Nitrogen contributes dark green color to plants, promote leaves, stems and other vegetative growth and improve fruit quality in plants<sup>57</sup>. PGPR in the primed seeds converts atmospheric nitrogen into plant absorbable form by the process of biological nitrogen fixation (BNF) due to the presence of nitrogenase which is coded by *nif* gene<sup>58,59</sup>. Glick *et al.*,<sup>60</sup> and Ahemad and Khan,<sup>61</sup> reported that nitrogen-fixing PGPR may be symbiotic like *Diazotrophs* (forms non-obligated interaction) and *Rhizobia* (forms nodules) and non-symbiotic like *Azospirillum*, *Azotobacter* and *Cyanobacteria* etc. A major portion of soil phosphorus is in insoluble

form<sup>62</sup>, phosphorus solubilising microorganism (PSM) like *Azotobacter*, *Bacillus*, *Beijerinckia*, *Enterobacter*, *Microbacterium*, *Pseudomonas* and *Serratia* release complex or mineral dissolved compounds like organic anions, protons, hydroxyl ions, carbon dioxide or liberation of extracellular enzymes namely phosphatase leads to solubilize the phosphorus and make it available to plant<sup>63,64</sup>. Phosphorous is an essential part of nucleic acids, phosphoproteins, phospholipids, energy-rich phosphate molecules and enzymes in plants. Phosphorous influences the lateral root morphology, root development, root branching and root to shoot ratio<sup>65,66</sup>. Some PGPR used in bio-priming have the ability to solubilize potassium from potassium-bearing minerals (Mica, illite and Orthoclase) by excretion of organic acids (citric acid, tartaric acid and oxalic acid) directly dissolving the rock potassium or chelate the silicon ion<sup>67</sup>. Potassium improves nitrogen use efficiency, involved in enzyme activation, stomatal activity, water and nutrient transport, transport of sugar and photosynthesis in the plant<sup>68,69,70,71</sup>. Siderophore producing rhizobacteria improves iron uptake in oats and *Arabidopsis*<sup>72,73</sup>.

Siderophore production by bio inoculant in primed seeds has suppressed the disease and improved plant growth<sup>74,75</sup>. It forms a stable complex with other heavy metals like Al, Cd, Cu, In, Pb and Zn which are an environmental concern<sup>76,77</sup>. Some of the PGPR like *Pseudomonas putida*, *Bacillus* sp. and *Enterobacter* sp. etc. are able to produce 1-Amino cyclopropane-1- carboxylate (ACC) deaminase under stress condition and showed reduction in ethylene content in plant whereby facilitated its growth and development<sup>78</sup>. Bio-priming of wheat with rhizobacteria containing ACC deaminase activity increased the growth and yield under stress condition<sup>79</sup>. Pattern and Glick<sup>80</sup>, reported that PGPR possesses the ability of synthesis and release of auxin which is an enhancer of plant cell division and

differentiation, xylem and root development, and initiation of lateral and adventitious root formation. Bio-priming of *Salvia officinalis* with PGPR having the ability to produce moderate auxin, recorded high germination rate, reduced speed of germination and higher root growth<sup>81</sup>. Glick<sup>64</sup>, reported bio-priming with IAA releasing rhizobacteria loosened plant cell walls and increased the root exudation from the plant which enhanced the availability of additional nutrients to the microorganism and supported its growth in the rhizosphere. The overproduction of ACC due to bio-priming with IAA producing microorganisms negatively regulated the root growth<sup>82</sup>. The gibberellin and cytokinin produced by *Azotobacter chroococcum* and *Rhizobium leguminosarum* have a positive effect on plant height and growth<sup>19,21</sup>.

#### Effect on physiological characters

Root is a key nutrient feeder to plant, further quality root biomass leads to better plant productivity. Root architecture (root length, root depth, root thickness and root to shoot ratio etc.) are playing an important role in strengthening the plant to survive in water deficit environment<sup>83</sup>. Seed bio-priming with PGPR reported to enhance the root related traits and producing a vigorous root system. Marcela et al.,<sup>84</sup> reported an increased root to leaves ratio in bio-primed rice. High root to shoot ratio, root length, root dry weight increased leaves number, leaf area and chlorophyll content have reported in bio-primed crops<sup>85,86</sup>. Anitha et al.,<sup>87</sup> reported increased root length and shoot length in soybean after bio-priming. Marcela et al.<sup>84</sup> reported increased lignin content in PGPR treated rice. Lignin involved in the better organization of macro fibrils which mediate the structural stability of the root cell wall<sup>88</sup>. Plant growth, nutrient uptake and nutrient use efficiency, synchronisation in germination and vigorous plant growth, speed of germination, and good plant stand under normal and stressed condition have been increased by seed bio-priming<sup>89,90,91,92</sup>. In bio-primed crops

increased biomass production grain yield and productivity were the evidence of growth stimulation<sup>89,93,94,95</sup>. Bio-priming is a better solution for enhanced growth and development of micro-propagated plants which have reduced photosynthetic activity, poorly functioning stomata and underdeveloped root and shoot system<sup>96</sup>. Bio-priming with PGPR minimizing the time required for lignification of micro-propagated plants and accelerates production process<sup>96,97</sup>.

#### Effects on biochemical parameters

In bio-priming, seeds are exposed to restricted water under controlled conditions which leads to the solute accumulation in the embryo and there will not be any germination till the embryo water potential reaches the threshold level required for radical emergence<sup>98</sup>. Bio-primed seeds have an advantage over non-primed seeds at the early process of germination because bio-primed seeds have large carbohydrate storage reserves which strengthen the plant to survive from low oxygen stress under flooded condition<sup>99</sup>.

Bio-priming leads to biochemical changes viz., enhanced production of proteins, hormones, phenol and flavonoid compounds contribute to better plant growth and development performance. Growth responses in herbaceous plants are determined by nitrogen reserve compounds like nitrates, amino acids and proteins<sup>100,101</sup>. Soluble protein percentage in bio-primed seeds and seedlings were higher compared to non-primed<sup>102</sup>. There was gain in total protein content and free amino acid content during the different growth stages after bio-priming with PGPR<sup>103,104,105</sup>. Soluble sugars act as osmolytes to maintain cell homeostasis, further by regulating the signals and acting as primary messenger it plays an important role in the expression of different genes responsible for plant growth and metabolism in source and sink tissues<sup>106,107,108</sup>. Total soluble sugar and reduced sugar content in plant increased after bio-priming<sup>109</sup>. Efficient mitochondrial development by augmenting energy metabolism i.e. high ATP

pool and an efficient ATP producing system occurs due to early imbibition process in primed seeds<sup>110</sup>. PGPR used in bio-priming enhanced the synthesis of specific phenolic acid in plants at different growth stages<sup>111</sup>. The combined application of PGPR increases the total phenol content in rice<sup>84</sup>. Increased in indole acetic acid content in the plant by bio-priming enhanced number of roots, root hairs and root area, plant cell division, cell differentiation, xylem and root development, lateral and adventitious root formation, pigment formation and biosynthesis of variable metabolites<sup>112</sup>. Besides plant growth promotion, bio-priming also stimulate the production of defense-related enzymes (peroxidase, superoxide dismutase, catalase, chitinase, ammonia lyases, etc.) which offer fitness benefit to plants against biotic and abiotic stress.

#### Effects on gene expression

Transcript level expression modulation is a key to a living system for response. Bio-priming leads to transcriptome level changes that make plant quick responsive. Microtubules consist of  $\alpha$ -tubulins and  $\beta$ -tubulins, which are the structural elements in cell growth and morphogenesis, and also involved in regulation and signal transduction<sup>113</sup>. Accumulation of  $\beta$ -tubulin preceded DNA replication and in primed seeds, the induction of DNA synthesis was started earlier, approximately 12 h earlier than untreated seeds. Amount of induced nuclear replication activity and  $\beta$ -tubulin was higher after priming in pepper and were found to be correlated with improved seed germination<sup>114</sup>. The transcriptome level studies provide a platform for comparative analysis of the plant roots response towards PGPR used in bio-priming and a pathogen. Repairing of the damaged part of the seeds and reduction in metabolic exudates were facilitated with priming by early transcription and protein synthesis<sup>115</sup>. A transcriptome level study of *Bacillus subtilis* was done by Xie *et al.*,<sup>116</sup> in

response to interaction with rice. According to that study, 176 genes showed a significant change in expression level in bacteria. Among these, 52 up-regulated genes were involved in metabolism and transport of nutrients and stress responsive where as 24 genes were down-regulated and involved in chemotaxis, motility, sporulation and teichuronic acid biosynthesis. Priming of rice plant with *Pseudomonas fluorescence* showed over expression of *RuBisCO*, led to increased photosynthesis activity<sup>117</sup>. Priming enhanced the expression of *RuBisCO* and *chl a/b* binding genes which increased carboxylation capacity and photosynthesis efficiency<sup>118,119</sup>. The respiration, energy metabolism and early reserve mobilization events in crops were regulated by bio-priming<sup>110,120</sup>. Up-regulation of expansin gene which is responsible for cell wall loosening was found important for coleoptile elongation<sup>121,122</sup>. The loosening of cell due to expansin occurs by disrupting hydrogen bonds between cellulose microfibrils and matrix polymers<sup>123</sup>. Enhanced expression of expansin genes was reported by Hussain *et al.*,<sup>118</sup> in bio-primed rice plant under submerged condition. Further, the enhanced expression of *NADH-GOGAT* and *GAPDH*, involved in energy production and biomass accumulation were reported in rice<sup>124,118</sup>. Putative glutathione s-transferase (*GST*) gene, an auxin-inducible gene which recognizes and transfers broad spectrum of reactive electrophilic compounds produced exogenously and endogenously to the vacuole by glutathione pump<sup>124,125</sup>. Kandasamy *et al.*,<sup>117</sup> found that overexpression of *GST* in rice plant primed with *Pseudomonas fluorescence* have an essential role in induced systemic resistance (ISR) and protecting cells from oxidative damage. Bio-priming also induced overexpression of genes involved in the regulation of secondary metabolism, development, transport protein and metal handling.

Table 1: Major groups of PGPR based on the application in agriculture

Group	Application	PGPR	Reference
Bio-fertilizer	Increasing the availability of nutrients to plant	<i>Azotobacter chroococcum</i>	Kumar et al., <sup>15</sup>
		<i>Gluconacetobacter diazotrophis</i>	Saravanan et al., <sup>16</sup>
		<i>Bacillus</i> sp.	Canbolat et al., <sup>17</sup>
		<i>Stenotrophomonas maltophilia</i>	Mehnaz et al., <sup>18</sup>
Phyto stimulator	Plant growth promotion generally through phyto hormones	<i>Rhizobium leguminosarum</i>	Noel et al., <sup>19</sup>
		<i>Bradyrhizobium</i> sp.	Antoun et al., <sup>20</sup>
		<i>Azotobacter chroococcum</i>	Verma et al., <sup>21</sup>
		<i>Xanthomonas</i> sp.	Sheng and Xia, <sup>22</sup>
		<i>Rhizobium phaseoli</i>	Zahir et al., <sup>23</sup>
Rhizo remediation	Degrading organic pollutant	<i>Cluyvera ascorbata</i>	Genrich et al., <sup>25</sup>
		<i>Pseudomonas putida</i>	Tripathi et al., <sup>26</sup>
		<i>Bradyrhizobium</i> sp.	Dary et al., <sup>27</sup>
		<i>Psychrobacter</i> sp.	Ma et al., <sup>28</sup>
Biopesticide	Controlling diseases mainly by producing antibiotics and antifungal metabolites	<i>Pseudomonas chlororaphis</i>	Liu et al., <sup>29</sup>
		<i>Bacillus subtilis</i>	Cazorla et al., <sup>30</sup>
		<i>Pseudomonas fluorescens</i>	Braud et al., <sup>31</sup>
		<i>Streptomyces</i> sp.	Bhattacharyya and Jha, <sup>32</sup>
		<i>Micromonospora</i> sp.	Franco-correa et al., <sup>33</sup>

## CONCLUSION

Bio-priming has a direct and indirect effect on plant growth and development. PGPR used in bio-priming have the ability to produce plant growth hormones (indole acetic acid, gibberellin and cytokinin), also have nutrient solubilization (phosphate, zinc and potassium) and nitrogen fixation ability which help plant for better growth and development. Further, bio-priming also act as a biocontrol agents producing some antibiotics, antifungal metabolites and by degrading organic pollutants. Bio-priming has reported a positive impact on seedling vigour, germination percentage, speed of germination, growth, development, yield and productivity of crops. Bio-priming with PGPR enhances the root related traits and producing vigorous plant root system, which is key for better production. Bio-priming also improve the total soluble sugar, reduced sugar content, total protein content and ATP production in plant that leads to optimum plant growth. Efficient mitochondrial development by augmenting energy metabolism is a speciality of bio-primed seed. Induction of DNA synthesis, transcription and protein synthesis starts

earlier in primed seeds than the non-primed seeds. Further, bio-priming found to enhance the expression of *RuBisCO*, *chl a/b* protein-coding genes, expansins genes, *GST* gene and  $\beta$ -tubulin gene whereby positively modulate the plant transcriptome for better growth, development and tolerance against biotic and abiotic stress. Bio-priming improves overall plant growth and development by physiological, biochemical and molecular level alteration which at the end resulted in to an asset to the modern agriculture.

## REFERENCES

1. Anonymous, Sector-wise contribution of GDP of India. Ministry of Statistics and Programme Implementation and Planning commission, Government of India. *Statistics Times*; <http://Statisticstimes.com/economy/sector-wise-gdp-contribution-of-India.php> (2017).
2. Walker, T. S., Bais, H. P., Grotewold, E. and Jorge M. V., Root exudation and rhizosphere biology. *Pl. Physiol.*, **132**: 44–51 (2003).

3. Kloepper, J. W., Leong, J., Teintze, M. and Schroth, M. N., Enhanced plant growth by siderophores produced by plant growth-promoting rhizobacteria. *Nature.*, **286**: 885–886 (1980).
4. Kavino, M., Harish, S., Kumar, N., Saravanakumar, D. and Samiyappan, R., Induction of systemic resistance in banana (*Musa* spp.) against *Banana bunchy topvirus* (BBTV) by combining chitin with root-colonizing *Pseudomonas fluorescens* strain CHA0. *Eur. J. Plant Pathol.*, **120**: 353-362 (2008).
5. Glick, B. R., The enhancement of plant growth by free-living bacteria. *Can. J. Microbiol.*, **41**: 109-117 (1995).
6. Patten, C. L. and Glick, B. R., Role of *Pseudomonas putida* indole acetic acid in development of the host plant root system. *Appl. Environ. Microbiol.*, **68**: 3795-3801 (2002).
7. Wang, C., Knill, E., Glick, B. R. and Defago, G., Effect of transferring 1-aminocyclopropane-1-carboxylic acid (ACC) deaminase genes into *Pseudomonas fluorescens* strain CHA0 and its gacA derivative CHA96 on their growth-promoting and disease-suppressive capacities. *Can. J. Microbiol.*, **46**: 898–907 (2000).
8. Ramamoorthy, V. and Samiyappan, R., induction of defense-related genes in *Pseudomonas fluorescent* treated chili plants in response to infection by *Colletotrichum capsic*. *J. Myco. Pl. Pathol.*, **31**: 146-155 (2001).
9. Kavino, M., Harish, S., Kumar, N., Saravanakumar, D., Damodaran, T., Soorinathasundaram, K. and Samiyappan, R., Rhizosphere and endophytic bacteria for induction of systemic resistance of banana plantlets against *Bunchy topvirus*. *Soil Biol. Biochem.*, **39**: 1087-1098 (2007).
10. Grichko, V. P. and Glick, B. R., Amelioration of flooding stress by ACC deaminase-containing plant growth-promoting bacteria. *Pl. Physiol. Biochem.*, **39**: 11–17 (2001).
11. Mayak, S., Tirosh, T. and Glick, B. R., Plant growth-promoting bacteria that confer resistance to water stress in tomato and pepper. *Pl. Sci.*, **166**: 525–530 (2004a).
12. Mayak, S., Tirosh, T. and Glick, B. R., Plant growth promoting bacteria that confer resistance in tomato and pepper to salt stress. *Pl. Physiol. Biochem.*, **167**: 650–656 (2004b).
13. Hemmatollah, P., Yaser, Y. and Valiollah, B., The effect of *Piriformospora indica* seed bio-priming and paclobutrazol foliar spraying on tolerance to chilling stress in green beans (*Phaseolus vulgaris* L.). *Environ. stress crop sci.*, **10(4)**: 459-474 (2017).
14. Jing, Y., He, Z. and Yang, X., Role of rhizobacteria in phytoremediation of heavy metal contaminated soil. *J. Zhejiang Univ. Sci. B.*, **8(3)**: 192-207 (2007).
15. Kumar, V., Behl, R. K. and Narula, N., Establishment of phosphate solubilizing strains of *Azotobacter chroococcum* in the rhizosphere and their effect on wheat cultivars under greenhouse conditions. *Microbiol. Res.*, **156**: 87–93 (2001).
16. Saravanan, V. S., Madhaiyan, M. and Thangaraju, M., Solubilization of zinc compounds by the diazotrophic, plant growth promoting bacterium *Gluconacetobacter diazotrophicus*. *Chemospher.*, **66**: 1794–1798 (2007).
17. Canbolat, M. J., Bilen, S. C., Akmakc, R., Sahin, F. and Aydin, A., Effect of plant growth-promoting bacteria and soil compaction on barley seedling growth, nutrient uptake, soil properties and rhizosphere microflora. *Biol. Fertil. Soil.*, **42**: 350–357 (2006).
18. Mehnaz, S., Baig, D. N. and Lazarovits, G., Genetic and phenotypic diversity of plant growth promoting rhizobacteria isolated from sugarcane plants growing in Pakistan. *J. Microbiol. Biotechnol.*, **20**: 1614–1623 (2010).
19. Noel, T. C., Sheng, C., Yost, C. K., Pharis, R. P. and Hynes, M. F., *Rhizobium leguminosarum* as a plant growth

- promoting rhizobacterium: direct growth promotion of canola and lettuce. *Can. J. Microbiol.*, **42**: 279–283 (1996).
20. Antoun, H., Beauchamp, C. J., Goussard, N., Chabot, R. and Lalonde, R., Potential of *Rhizobium* and *Bradyrhizobium* species as plant growth promoting rhizobacteria on non-legumes: effects on radishes (*Raphanus sativus* L.). *Plant Soil.*, **204**: 57–67 (1998).
  21. Verma, A., Kukreja, K., Pathak, D. V., Suneja, S. and Narula, N., In vitro production of plant growth regulators (PGRs) by *Azorobacter chroococcum*. *Ind. J. Microbiol.*, **41**: 305–307 (2001).
  22. Sheng, X. F. and Xia, J. J., Improvement of rape (*Brassica napus*) plant growth and cadmium uptake by cadmium-resistant bacteria. *Chemosphere*, **64**: 1036–1042 (2006).
  23. Zahir, Z. A., Shah, M. K., Naveed, M. and Akhter, M. J., Substrate dependent auxin production by *Rhizobium phaseoli* improves the growth and yield of *Vigna radiata* L. under salt stress conditions. *J. Microbiol. Biotechnol.*, **20**: 1288–1294 (2010).
  24. Wani, P. A. and Khan, M. S., *Bacillus* species enhance growth parameters of chickpea (*Cicer arietinum* L.) in chromium stressed soils. *Food Chem. Toxicol.*, **48**: 3262–3267 (2010).
  25. Genrich, I. B., Dixon, D. G. and Glick, B. R., A plant growth promoting bacterium that decreases nickel toxicity in seedlings. *Appl. Environ. Microbiol.*, **64**: 3663–3668 (1998).
  26. Tripathi, M., Munot, H. P., Shouch, Y., Meyer, J. M. and Goel, R., Isolation and functional characterization of siderophore-producing lead- and cadmium-resistant *Pseudomonas putida* KNP9. *Curr. Microbiol.*, **5**: 233–237 (2005).
  27. Dary, M., Chamber-Perez, M. A., Palomares, A. J. and Pajuelo, E., “In situ” phyto stabilisation of heavy metal polluted soils using *Lupinus luteus* inoculated with metal resistant plant-growth promoting rhizobacteria. *J. Hazard. Mater.*, **177**: 323–330 (2010).
  28. Ma, Y., Rajkumar, M., Vicente, J. A. and Freitas, H., Inoculation of Ni-resistant plant growth promoting bacterium *Psychrobacter sp.* strain SRS8 for the improvement of nickel phytoextraction by energy crops. *Int. J. Phytorem.*, **13**: 126–139 (2011).
  29. Liu, H., He, Y., Jiang, H., Peng, H., Huang, X., Zhang, X., Thomashow, L. S. and Xu, Y., Characterization of a phenazine producing strain *Pseudomonas chlororaphis* GP72 with broad-spectrum antifungal activity from green pepper rhizosphere. *Curr. Microbiol.*, **54**: 302–306 (2007).
  30. Cazorla, F. M., Romero, D., Perez-García, A., Lugtenberg, B. J. J., de Vicente, A. and Bloemberg, G., Isolation and characterization of antagonistic *Bacillus subtilis* strains from the avocado rhizosphere displaying bio control activity. *J. Appl. Microbiol.*, **103**: 1950–1959 (2007).
  31. Braud, A., Jezequel, K., Bazot, S. and Lebeau, T., Enhanced phyto extraction of an agricultural Cr-, Hg- and Pb-contaminated soil by bio-augmentation with siderophore-producing bacteria. *Chemosphere*, **74**: 280–286 (2009).
  32. Bhattacharyya, P. N. and Jha, D. K., Plant growth-promoting rhizobacteria (PGPR): emergence in agriculture. *World J. Microbiol. Biotechnol.*, **28**: 1327–1350 (2012).
  33. Franco-Correa, M., Quintana, A., Duque, C., Suarez, C., Rodríguez, M. X., and Barea, J. M., Evaluation of *Actinomyces* strains for key traits related to plant growth promotion and mycorrhiza helping activities. *Appl. Soil Ecol.*, **45**: 209–217 (2010).
  34. Abhilash, P. C., Tripathi, V., Edrisi, S. A., Dubey, R. K., Bakshi, M., Dubey, P. K., Singh, H. B. and Stephen, D. E., Sustainability of crop production from polluted lands. *Energ. Ecol. Environ.*, **1(1)**: 54–65 (2016).
  35. Rainer, B., Use of plant-associated *Bacillus* strains as biofertilizers and

- biocontrol agents in agriculture. Dinesh, K. M. (Ed.). Bacteria in Agrobiolgy: Plant growth responses. Springer Heidelberg Dordrecht London, New York. DOI 10.1007/978-3-642-20332-9 (2011).
36. Staley, T. E. and Drahos, D. J., Marking soil bacteria with lacZY. In: Weaver, R. W., Angel, J. S., Bottomley, P. J. (Eds). Methods of Soil Analysis. Part 2. Microbiological and Biochemical Properties. Soil Science Society of America, Madison, WI, pp: 689–706 (1994).
37. Gindrat, D., *Alternaria radicina*, an important parasite of market garden Umbelliferae. *Rev. Suisse Vitic. Arboric. Horti.*, **11**: 257–267 (1979).
38. Bashan Y., Inoculants for plant growth-promoting bacteria in agriculture. *Biotechnol. Adv.*, **16**: 729–70 (1998).
39. Srinivasan, K., Gilardi, G., Garibaldi, A. and Gullino, M.L., Bacterial antagonists from used rockwool soilless substrates suppress *Fusarium* wilt of tomato. *J. Plant Pathol.*, **91**: 147–54 (2009).
40. Munif, A., Hallmann, J. and Sikora, R. A., The influence of endophytic bacteria on *Meloidogyne incognita* infection and tomato plant growth, *J. ISSAAS.*, **19**: 68–74 (2013).
41. Bashan, Y. and Levanony, H., Current status of *Azospirillum* inoculation technology: *Azospirillum* as a challenge for agriculture, *Can. J. Microbiol.*, **36**: 591–608 (1990).
42. Duarte, C. R., Neto, J. L. V., Lisboa, M. H., Santana, R. C., Barrozo, M. A. S. and Murata, V. V., Experimental study and simulation of mass distribution of the covering layer of soybean seeds coated in a spouted bed. *Braz. J. Chem. Eng.*, **21**: 59–67 (2004).
43. Heydecker, W., Germination of an idea: the priming of seeds. *University of Nottingham School of Agriculture Report* 50–67 (1973).
44. Bradford, K. J., Manipulation of seed water relation via osmotic priming to improve germination under stress condition. *Hort. Sci.*, **21**: 1105–1112 (1986).
45. Mabhaudhi, T. and Modi, A. T., Can hydro-priming improve germination vigor, speed and emergence of maize landraces under water stress? *J. Agri. Sci. Tech. B.*, **1**: 20–28 (2011).
46. Golezani, G. K., Jabbarpour, S., Zehtab-Salmasi, S. and Mohammadi, A., Response of winter rapeseed (*Brassica napus* L.) cultivars to salt priming of seeds. *Afr. J. Agric.*, **5**: 1089-1094 (2010).
47. Soughir, M., Elouaer, M. A. and Cherif, H., Effects of NaCl priming duration and concentration on germination behaviour of fenugreek. *Albanian J. Agric. Sci.*, **11**: 193-198 (2012).
48. Ashraf, M. and Rauf, H., Inducing salt tolerance in maize (*Zea mays* L.) through seed priming with chloride salts: Growth and ion transport at early growth stages. *Acta. Physiol. Pl.*, **23**: 407–414 (2001).
49. Afzal, S., Nadeem, A., Zahoor, A. and Qaiser, M., Role of seed priming with zinc in improving the hybrid maize (*Zea mays*) yield. *American-Eurasian. J. Agric. Environ. Sci.*, **13(3)**: 301-306 (2006).
50. Larissa, C. S., Marina, A. G., Marcelo, L. C., Victor, D. A. and Rogerio, F. C., Effects of hormonal priming on seed germination of pigeon pea under cadmium Stress. *An. Acad. Bras. Ciênc.*, **87(3)**: 1847-1852 (2015).
51. Arin, L. and Kiyak, Y., The effects of pre-sowing treatments on emergence and seedling growth of tomato seed (*Lycopersicon esculentum* Mill.) under several stress conditions. *Pak. J. Biol. Sci.*, **6**: 990-994 (2003).
52. Golezani, G. K., Aliloo, A. A., Valizadeh, M. and Moghaddam, M., Effects of hydro and osmopriming on seed germination and field emergence of Lentil (*Lens culinaris* Medik.). *Bot. Hort. Agro. Bot. Cluj.* **36**: 29-33 (2008).
53. Navin, P., Pravin, P., Shailesh, K. T., Manimurugan, C., Sharma, R. P. and Singh, P. M., Osmo-priming of tomato genotypes with polyethylene glycol 6000

- induces tolerance to salinity stress. *Trends Biosci.*, **7(24)**: 4412-4417 (2014).
54. Nithya, N., Geetha, R. and Prakasam, V., Alleviation of the adverse effects of salinity stress in rice var.PMK 4 (*Oryza sativa* L.) by seed bio-priming with salinity tolerant *Pseudomonas fluorescent*. *Ann. Pl. Soil Res.*, **17**: 190-193 (2015).
55. Suresh Rao, K. S., Pradeep Rao, T. K., Dnyanobarao, G. S., Agrawal, T. and Kotasthane, A. S., Root growth stimulation in rice (*Oryza sativa* L.) by seed bio-priming with *Trichoderma* sp. *Appl. Biol. Res.*, **18(1)**: 30-38 (2016).
56. Shah, J. L., Niaz, A. W., Ghulam, M. L., Abdul, H. L., Ghulam, M. B., Khalid, H. T., Tofique, A. B., Safdar, A. W. and Ayaz, A. L., Role of nitrogen for plant growth and development: A review. *Adv. Environ. Biol.*, **10(9)**: 209-218 (2016).
57. Bloom, A. J., Photorespiration and nitrate assimilation: major intersection between plant carbon and nitrogen. *Photosynth. Res.*, **123**: 117- 128 (2015).
58. Thakuria, D., Talukdar, N. C., Goswami, C., Hazarika, S., Boro, R. C. and Khan, M. R., Characterization and screening of bacteria from rhizosphere of rice grown in acidic soils of Assam. *Curr. Sci.*, **86**: 978–985 (2004).
59. Kim, J. and Rees, D. C., Nitrogenase and biological nitrogen fixation. *Biochem.*, **33**: 389–397 (1994).
60. Glick, B. R., Patten, C. L., Holguin, G. and Penrose, G. M., Biochemical and genetic mechanisms used by plant growth promoting bacteria. Imperial College Press, London (1999).
61. Ahemad, M. and Khan, M. S., Evaluation of plant growth promoting activities of rhizobacterium *Pseudomonas putida* under herbicide-stress. *Ann. Microbiol.*, **62**: 1531–1540 (2012).
62. Khan, M. S., Zaidi, A., Wani, P. A. and Oves, M., Role of plant growth promoting rhizobacteria in the remediation of metal contaminated soils. *Environ. Chem. Lett.*, **7**: 1–19 (2009).
63. Zaidi, A., Khan, M. S., Ahemad, M. and Oves, M., Plant growth promotion by phosphate solubilizing bacteria. *Acta. Microbiol. Immunol. Hung.* **56**: 263–284 (2009).
64. Glick, B. R., Plant Growth-Promoting Bacteria: Mechanisms and Applications. *Hindawi Publishing Corporation, Scientifica* (2012).
65. Lopez-Bucio, J., Hernandez-Abreu, E., Sanchez-Calderon, L., Nieto- Jacobo, M. F., Simpson, J. and Herrera-Estrella, L., Phosphate availability alters architecture and causes changes in hormone sensitivity in the *Arabidopsis* root system. *Pl. Physiol.*, **129**: 244-256 (2002).
66. Jin, J., Wang, G. H., Liu, X. B., Pan, X. W. and Herbert, S. J., Phosphorus application affects the soybean root response to water deficit at the initial flowering and full pod stages. *Soil Sci. Plant Nut.*, **51**: 953–960 (2005).
67. Sheng, X. F. and He, L. Y., Solubilization of potassium-bearing minerals by a wild-type strain of *Bacillus edaphicus* and its mutants and increased potassium uptake by wheat. *Can. J. Microbiol.*, **52**: 66-72 (2006).
68. Prajapati, K. B. and Modi, H. A., The importance of potassium in plant growth – A Review. *Ind. J. Pl. Sci.*, **1**: 177-186 (2012).
69. Van Brunt, J. M. and Sultenfuss, J. H., Better crops with plant food. *In Potassium: Functions of Potassium*, **82(3)**: 4-5 (1998).
70. Thomas, T. C. and Thomas, A. C., Vital role of potassium in the osmotic mechanism of stomata aperture modulation and its link with potassium deficiency. *Pl. Signal. Behav.*, **4(3)**: 240–243 (2009).
71. Schwartzkopf, C., *Potassium, calcium, magnesium- how they relate to plant growth* mid-continent agronomist, us green section role of potassium in crop establishment from agronomists of the potash & phosphate institute (1972).

72. Crowley, D. E. and Kraemer, S. M., Function of siderophores in the plant rhizosphere. In: Pinton, R., Varanini, Z. and Nannipieri, P., (Eds.). The rhizosphere: Biochemistry and Organic substances at the soil-plant interface, Second CRC Press, pp: 173–200 (2007).
73. Vansuyt, G., Robin, A., Briat, J. F., Curie, C. and Lemanceau, P., Iron acquisition from Fe-pyoverdine by *Arabidopsis thaliana*. *Mol. Pl. Microbe Interact.*, **20**: 441–447 (2007).
74. Keswani, C., Mishra, S., Sarma, B. K., Singh, S. P. and Singh, H. B., Unraveling the efficient application of secondary metabolites of various *Trichoderma*. *Appl. Microbiol. Biotechnol.*, **98**: 533-544 (2014).
75. Jain, A., Singh, S., Sarma, K. B. and Singh, H. B., Microbial consortium-mediated reprogramming of defence network in pea to enhance tolerance against *Sclerotinia sclerotiorum*. *J. Appl. Microbiol.*, **112**: 537- 550 (2012).
76. Kiss, T. and Farkas, E., Metal-binding ability of desferrioxamine. *B. J. Inclusion Phenom. Mol. Recognit. Chem.*, **32**: 385–403 (1998).
77. Neubauer, U., Furrer, G., Kayser, A. and Schulin, R., Siderophores, NTA, and citrate: potential soil amendments to enhance heavy metal mobility in phytoremediation. *Int. J. Phytorem.*, **2**: 353–368 (2000).
78. Saleem, M., Arshad, M., Hussain, S. and Bhatti, A. S., Perspective of plant growth promoting rhizobacteria (PGPR) containing ACC deaminase in stress agriculture. *J. Ind. Microbiol. Biotechnol.*, **34**: 635–648 (2007).
79. Zia-ul-hassan, Ansari, T. S., Shah, A. N., Jamro, G. M. and Rajpar, I., Bio-priming of wheat seeds with rhizobacteria containing ACC-deaminase and phosphate solubilizing activities increases wheat growth and yield under phosphorus deficiency. *Pak. J. Agri., Agril. Engg., Vet. Sci.*, **31(1)**: 24-32 (2015).
80. Patten, C. L. and Glick, B. R., Bacterial biosynthesis of indole-3-acetic acid. *Can. J. Microbiol.*, **42**: 207–220 (1996).
81. Mansour, G. and Mehrnaz, H., Bio-priming of *Salvia officinalis* seed with growth promoting rhizobacteria affects invigoration and germination indices. *J. Biol. Environ. Sci.* **8(22)**: 29-36 (2014).
82. Glick, B. R., Penrose, D. M. and Li, J. P., A model for the lowering of plant ethylene concentrations by plant growth-promoting bacteria. *J. Theor. Biol.*, **190(1)**: 63-68 (1998).
83. O'Toole, J. C. and Bland, W. L., Genotypic variation in crop plant root systems. *Adv. Agron.*, **41**: 91-145 (1987).
84. Marcela, C. F., Fernanda, I. B., Marta, C. C., Letícia, A. G. and Gisele, B. S., Morpho-anatomical and biochemical changes in the roots of rice plants induced by plant growth promoting microorganisms. *J. Bot.*, **14**: 144-147 (2014).
85. Priya, P., Patil, V. C. and Kumar, B. A., Characterization of grain sorghum (*Sorghum bicolor* L.) for root traits associated with drought tolerance. *Res. Environ. Life Sci.*, **9(2)**: 163–165 (2016).
86. Rawat, L., Singh, Y., Shukla, N. and Kumar, J., Seed bio-priming with salinity tolerant isolates of *Trichoderma harzianum* alleviates salt stress in rice: growth, physiological and biochemical characteristics. *J. Pl. Pathol.*, **94(2)**: 353-365 (2012).
87. Anitha, U. V., Gatti, M. and Jahagirdar, S., Influence of seed priming agents on yield, yield parameters and purple seed stain disease in soybean. *Karnataka J. Agric. Sci.*, **28(1)**: 20-23 (2015).
88. Rubin, E. M., Genomics of cellulosic biofuels. *Nature*, **454(7206)**: 841–845 (2008).
89. Yadav, S. K., Dave, A., Sarkar, A., Singh, H. B. and Sarma, B. K., Co-inoculated bio-priming with *Trichoderma*, *Pseudomonas* and *Rhizobium* improves crop growth in *Cicer arietinum* and *Phaseolus vulgaris*. *Inter. J. Agric.*

- Environ. Biotechnol.*, **6(2)**: 255–259 (2013).
90. Tanwar, A., Aggarwal, A., Kaushish, S. and Chauhan, S., Interactive effect of AM fungi with *Trichoderma viride* and *Pseudomonas fluorescens* on growth and yield of broccoli. *Pl. Prot. Sci.*, **49**: 137–145 (2013).
91. Moeinzadeh, A., Sharif-Zadeh, F., Ahmadzadeh, M. and Tajabadi, F. H., Bio-priming of sunflower (*Helianthus annuus* L.) seed with *Pseudomonas fluorescens* for improvement of seed invigoration and seedling growth. *Aust. J. Crop Sci.*, **4**: 564 (2010).
92. Muruli, C. N., Studies on seed priming and primed seed longevity in Onion (*Allium cepa* L.) and Capsicum (*Capsicum annuum* L.). M. Sc. (Ag.) Thesis, University of Agricultural Sciences, Bangalore (2013).
93. Karthika, C. and Vanangamudi, K., Bio-priming of maize hybrid COH (M) 5 seed with liquid biofertilizers for enhanced germination and vigor. *Acad. J.*, **8(25)**: 3310-3317 (2013).
94. Baral, B. R. and Adhikari, P., Effect of *Azotobacter* on growth and yield of maize. *SAARC J. Agri.*, **11(2)**: 141–147 (2013).
95. Namvar, A. and Khandan, T., Bio-priming and mineral fertilizers effects on agronomical performance of rapeseed (*Brassica napus* L.). *Ekologija*, **60(3)**: 54–63 (2014).
96. Kavino, M., Harish, S., Kumar, N., Saravanakumar, D. and Samiyappan, R., Effect of chitinolytic PGPR on growth, yield and physiological attributes of banana (*Musa* spp.) under field conditions. *Appl. soil Ecol.*, **45**: 71-77 (2010).
97. Ramamoorthy, V., Raguchander, T. and Samiyappan, R., Induction of defense-related proteins in tomato roots treated with *Pseudomonas fluorescent* Pfl and *Fusarium oxysporum* f. sp *lycopersici*. *Plant-soil*, **239**: 55-68 (2002).
98. Bradford, K. J., Manipulation of seed water relation via osmotic priming to improve germination under stress condition. *Hort. Sci.*, **21**: 1105–1112 (1986).
99. Ella, E. S., Dionisio-Sese, M. L. and Ismail, A. M., Seed pre-treatment in rice reduces damage, enhances carbohydrate mobilization and improves emergence and seedling establishment under flooded conditions. *AoB Plants: plr00*.doi: 10.1093/aobpla/plr007 (2011).
100. Cyr, D. R., Bewley, J. D. and Dumbroff, E. B., Seasonal variation in nitrogen storage reserves in the roots of leafy spurge (*Euphorbia escula*) and response to decapitation and defoliation, *Physiol. Plant.*, **78**: 361-366 (1990).
101. Volenec, J. J., Ourry, A. and Joern, B. C., A role for nitrogen reserves in forage regrowth and stress tolerance. *Physiol. Plant.*, **97**: 185-193 (1996).
102. Dhanya, B. A., Evaluation of microbial seed priming in relation to seed germination, plant growth promotion in *Morinda citrifolia* L. (Noni). M. Sc. (Ag.) Thesis. University of Agricultural Sciences, Bangalore (2014).
103. Aishwath, O. P., Lal, G., Kant, K., Sharma, Y. K., Ali, S. F. and Naimuddin., Influence of biofertilizers on growth and yield of coriander under typic haplustepts. *Inter. J. Seed Spic.*, **2**: 9-14 (2012).
104. Warwate, S. I., Kandoliya, U. K., Bhadja, N. V. and Golakiya, B. A., The effect of plant growth promoting rhizobacteria (PGPR) on biochemical parameters of coriander (*Coriandrum sativum* L.) seedling. *Int. J. Curr. Microbiol. App. Sci.*, **6(3)**: 1935-1944 (2017).
105. Ahmed, R. S., Mohamed, S. A., Abd, M. A. and Khalid, A., Potential impacts of seed bacterization or salix extract in faba bean for enhancing protection against bean yellow mosaic disease. *Nature and Sci.*, **12(10)**: 213-215 (2014).
106. Gupta, A. K. and Kaur, N., Sugar signaling and gene expression in relation to carbohydrate metabolism under abiotic stresses in plants. *J. Bio. Sci.*, **30**: 761-776 (2005).
107. Rolland, F., Baena-Gonzalez, E. and Sheen, J., Sugar sensing and signaling in

- plants: conserved and novel mechanisms. *Annu. Rev. Pl. Biol.*, **57**: 675-709 (2006).
108. Chen, J. G., Sweet sensor, surprising partners. *Sci. STKE*. **373**: 7 (2007).
109. Hafsa, N. and Asghari, B., Role of plant growth promoting rhizobacteria and their exopolysaccharide in drought tolerance of maize. *J. Pl. Inter.*, **9**: 689–701 (2014).
110. Chen, K. and Arora, R., Priming memory invokes seed stress-tolerance. *Environ. Exp. Bot.*, **94**: 33–45 (2013).
111. Singh, U. P., Sarma, B. K. and Singh, D. P., Effect of plant growth promoting rhizobacteria and culture filtrate of *Sclerotium rolfsii* on phenolic and salicylic acid contents in chickpea (*Cicer arietinum*). *Curr. Microbiol.*, **46**: 131-140 (2003).
112. Spaepen, S. and Vanderleyden, J., Auxin and plant-microbe interactions. *Cold Spring Harb. Perspect. Biol*, **3(4)**: a001438 (2011).
113. Mohammed R. F., Patrick, J. and Gulick., The  $\alpha$ -tubulin gene family in wheat (*Triticum aestivum* L.) and differential gene expression during cold acclimation. *Genome*, **50(5)**: 502-510 (2007).
114. Lanteri, S., Kraak, H. L., Ric De Vos, C. H., Bino, R. J., Effects of osmotic preconditioning on nuclear replication activity in seeds of pepper (*Capsicum annum* L.). *Physiologia Plantarum*, **89**: 433–440 (1993).
115. Entesari, I. M., Sharifzadeh, F., Ahmadzadeh, M. and Farhangfar, M., Seed bio-priming with *Trichoderma* sp. and *Pseudomonas fluorescence* growth parameters, enzymes activity and nutritional status of Soybean. *Int. J. Agron. Pl. Prod.*, **4(4)**: 610-619 (2013).
116. Xie, S., Wu, H., Chen, L., Zang, H., Xie Y. and Gao, X., Transcriptome profiling of *Bacillus subtilis* OKB105 in response to rice seedlings. *BMC Microbiol.* **15**: 21 (2015).
117. Kandasamy, S., Loganathan, K., Muthuraj, R., Duraisamy, S. and Seetharaman, S., Understanding the molecular basis of plant growth promotional effect of *Pseudomonas fluorescens* on rice through protein profiling. *Proteome Sci.*, **7**: 47 (2009).
118. Hussain, S., Yin, H., Peng, S., Khan, F. A., Khan, F., Sameeullah, M., Hussain, H. A., Huang, J., Cui, K. and Nie, L., Comparative transcriptional profiling of primed and non-primed rice seedlings under submergence stress. *Front. Plant Sci.*, **7**: 1125 (2016).
119. Mommer, L., Pons, T. L., Wolters-Arts, M., Venema, J. H. and Visser, E. J. W., Submergence-induced morphological, anatomical and biochemical responses in a terrestrial species affect gas diffusion resistance and photosynthetic performance. *Pl. Physiol.*, **139**: 497–508 (2005).
120. Paparella, S., Arau, J. S. S., Rossi, G., Wijayasinghe, M., Carbonera, D. and Balestrazzi, A., Seed priming: state of the art and new perspectives. *Pl. Cell Rep.*, **34**: 1281–1293 (2015).
121. Choi, D., Lee, Y., Cho, H. T. and Kende, H., Regulation of expansin gene expression affects growth and development in transgenic rice plants. *Plant Cell*, **15**: 1386–1398 (2003).
122. Magneschi, L. and Perata, P., Rice germination and seedling growth in the absence of oxygen. *Ann. Bot.*, **103**: 181–196 (2009).
123. McQueen-Mason, S. J. and Cosgrove, J. D., Disruption of hydrogen bonding between plant cell wall polymers by proteins that induce wall extension. *Proc. Nati. Acad. Sci.*, **91(14)**: 6574-657 (1994).
124. Tamura, W., Kojima, S., Toyokawa, A., Watanabe, H., Tabuchi-Kobayashi, M. and Hayakawa, T., Disruption of a novel *NADH*-glutamatesynthase2 gene caused marked reduction in spikelet number of rice. *Front. Pl. Sci.* **2**: 57 (2011).
125. Marrs, K. A., The function and regulation of glutathione S- transferases in plants. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, **47**: 127-157 (1996).
126. Ishikawa, T., The ATP-dependent glutathione S-conjugate export pump. *Trends Biochem. Sci.*, **17(11)**: 463-468 (1992).